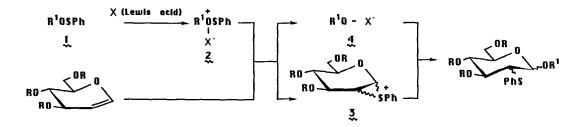
SULFENATE ESTERS AS GLYCOSYL ACCEPTORS: A NOVEL APPROACH TO O-GLYCOSIDES FROM THIOGLYCOSIDES AND SULFENATE ESTERS¹)

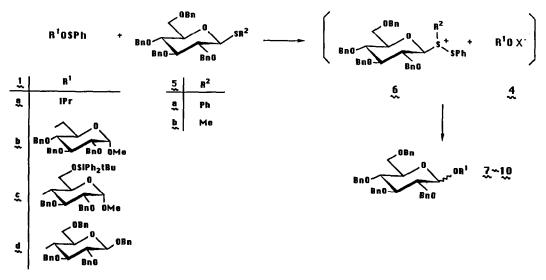
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Abstract: The reactions of thioglycosides and sulfenate esters were effected in the presence of Lewis acid to give corresponding O-glycosides. The stereoselectivity was highly dependent on the solvent.

Recently we reported a conceptually novel approach to the synthesis of 2-deoxyglycosides based on the Lewis acid mediated activation of sulfenate esters 1 readily obtainable from

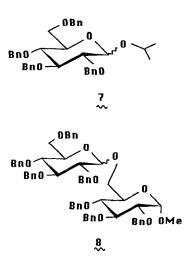


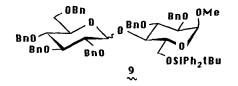
alcohols²). A sulfenate ester is activated by a Lewis acid to genarate an ionic species 2 and a glycal attacks as a nucleophile on a sulfur atom of 2 giving an episulfonium ion 3. It can be viewed that the reaction is a result of successive hard-hard (i.e. a Lewis acid and 1) and softsoft (i.e. 2 and a glycal) acid-base interactions. In recent years, potential utility of thioglycosides as glycosyl donors has been demonstrated in several reports³⁾⁻¹¹, most of

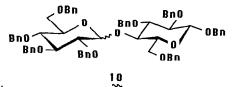


which take advantage of the specific affinity of sulfide groups to soft electrophiles. From these facts, it was assumed that an activated form 2 of a sulfenate ester would react with a thioglycoside 5 to form a cationic intermediate 6, which in turn reacts with a nucleophilic species 4 to give a glycosylated product.

As expected, thioglycosides $5a^{3(5)}$ and $5b^{1(2)}$ reacted smoothly with isopropyl benzenesulfenate $1a^{14}$ in the presence of a catalytic amount of TMSOTf and the product 7 was obtained in a high yield (table 1, entry 1~6). Sulfenate esters 1b, 1c and 1d derived from glucopyranosides also served satisfactorily as glycosyl acceptors to give disaccharides 8,9 and 10 (entry 7~12). All reactions proceeded under extremely mild conditions; less than 1h at -35°C in most of the cases. So far as 1-deoxy-1-thio-β-D-glucopyranoside 5b was investigated as a glycosyl donor, the stereoselectivity was solventdependent, being α -selective in diethyl ether and β -selective in acetonitrile¹⁵). The β -selectivity was higher in the sterically less hindered substrate (entry 8) and enhanced by lowering the temperature (entry 5, 6). On the contrary, the α -selectivity was higher in the more hindered substrate (entry 9) and scarecely affected by the temperature (entry 3, 4).







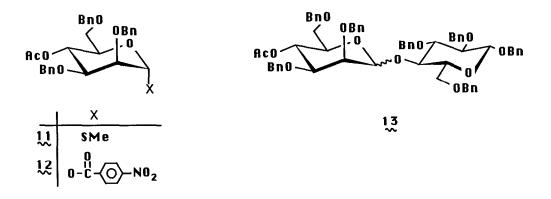
entry	sulfenate 1 ^{c)}	thioglycoside 5	molar ratio 5/1/TMSOTf	solvent	temp.(°C)/time	product	yield (%)	α:β
] a)	a	a	1/1.3/0.1	(CICH ₂) ₂	20/30min	7	70	44:56 ^d)
2a)	а	b	1/1.3/0.1	(CICH ₂) ₂	0/15min	7	93	44:56d)
3a)	а	Ь	1/1.3/0.1	Et ₂ O	0/15min	7	87	79:21d)
4a)	а	b	1/1.3/0.1	Et ₂ O	-35/30min	7	90	81:19 ^{d)}
5b)	а	b	1/1.3/0.1	MeCN	0/15min	7	88	21 :79 ^{d)}
6 ^b)	а	b	1/1.3/0.1	MeCN	-35/30min	7	89	12:88 ^d)
7a)	b	b	1/1.3/0.1	Et ₂ O	-35/40min	817)	65	67 :33ª)
8b) 9a)	b c	b b	1/1.3/0.1 1/1.3/0.2	MeCN Et ₂ O	-35/40min -35/20min	817) 918)	90 69	5 :95d) 86 :14e)
10b) 11a)	c d	b b	1/1.3/0.5 1/1.5/0.4	MeCN Et ₂ O	-35~20/18h -35/40min	918) 1019)	50 74	22 :78e) 66 :34d)
12 ^{b)}	d	b	1/1.5/0.3	MeCN	-35/30min	1019)	68	21 :79d)

Table 1Reactions of sulfenate esters with thioglycosides.

a) Carried out in the presence of molecular sieves 4A. b) Carried out in the presence of molecular sieves 3A. c) **1b**, **1c** and **1d** were prepared from corresponding alcohols (PhSCl, Et₃N, ether, -78°C) and used without purification. d) Determined by 400 MHz ¹H-NMR analysis. e) determined by individual isomer separation.

A typical glycosylation procedure was as follows (entry 8). A mixture of 5b (59.2 mg, 104 μ mol), 1b (77.5 mg, 135 μ mol), molecular sieves 4A (0.35 g) and acetonitrile (4 ml) was stirred at -35°C under dry nitrogen. TMSOTf (2 μ l, 10 μ mol) was added and the mixture was stirred at -35°C for 40 min. Triethylamine (60 μ l) was then added and the mixture was diluted with ethyl acetate (30 ml) and filtered through Celite. The filtrate was washed successively with aq. NaHCO₃ (30 ml) and brine (30 ml), dried over MgSO₄ and concentrated. Residue was purified by silca gel chromatography in 4:1 n-hexane-ethyl acetate to give 8 (92.6 mg, 90%; α : β =5:95), from which pure β -isomer was isolated by recrystallization from n-hexane-chloroform, m.p. 130~131.5°, $[\alpha]_D$ +18.4° (c 0.76)¹⁶) (lit.¹⁷) $[\alpha]_D$ +17.1° (c 0.42)).

As shown below, the present method could also be applicable to thiomannoside 11^{20} . Thus, 11 was reacted with sulfenate ester 1d to give a 67% yield of the disaccharide 13^{21}) which constitutes a structural unit of plant glycolipids (table 2, entry 1). Although the β -glycoside could not be obtained as a major product under the several conditions we attempted, a high α -selectivity was observed especially in diethyl ether.



		product 13				
entrya)	catalyst	11/1d/catalyst	solvent	temp.(°C)/time	yield(%)	α : β ^{b)}
1	TMSOTf	1/1.2/0.4	Et ₂ O	-35/30min	67	92:8
2	TMSOTf	1/1.6/0.4	MeCN	-35/30min	65	74:26
3	TrBF ₄	1/1.3/0.5	MeCN	-35/30min	44	77:23
4	BF ₃ •OEt ₂	1/1.5/0.5	MeCN	-35/30min	34	75:25

Table 2 Reactions of sulfenate ester 1d with thiomannoside 11

a) Carried out in the presence of molecular sieves 4A (entry 1) or 3A (entry 2~4).

b) Determined by individual isomer separation.

In conclusion, Lewis acid catalysed reaction of sulfenate esters with thioglycosides smoothly gave glycosylated products under mild conditions.

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References and Notes

- 1) Part of a series on "Novel approaches to glycoside synthesis". For the previous papers, see Ref. 2).
- 2) Y. Ito and T. Ogawa, Tetrahedron Lett., in press.
- 3) R. J. Ferrier, R. W. Hay and N. Vethaviyasar, Carbohydr. Res., 27, 55 (1973).
- 4) T. Mukaiyama, T. Nakatsuka and S. Shoda, Chem. Lett., 1979, 487.
- 5) J. W. Van Cleve, Carbohydr. Res., 70, 161 (1979).
- 6) S. Hanessian, C. Bacquet and N. Lehong, Carbohydr. Res., 80, C17 (1980).
- 7) P. J. Garegg, C. Henrichson and T. Norberg, Carbohydr. Res., 116, 162 (1983).
- 8) K. C. Nicolaou, S. P. Seitz and D. P. Papahatjis, J. Am. Chem. Soc., 105, 2430 (1983).
- 9) H. Lönn, Carbohydr. Res., 139, 105, 115 (1985).
- 10) a) P. Fügedi and P. J. Garegg, Carbohydr. Res., 149, C9 (1986).
 b) F. Andersson, P. Fügedi, P. J. Garegg and M. Nashed, Tetrahedron Lett., 27, 3919 (1986).
- 11) S. Sato, M. Mori, Y. Ito and T. Ogawa, Carbohydr. Res., 155, C6 (1986).
- 12) Prepared from corresponding tetraacetate¹³⁾ (1. NaOMe/MeOH, 2. PhCH₂Br, NaH/DMF). m.p. 64-66°C, $[\alpha]_D$ +12.0° (c 2.3).
- 13) T. Ogawa and M. Matsui, Carbohydr. Res., 54, C17 (1977).
- 14) L. L. Chang, D. B. Denney, D. Z. Denney and R. J. Kazior, J. Am. Chem. Soc., 99, 2293 (1977).
- 15) For similar solvent effect, see: S. Hashimoto, M. Hayashi and R. Noyori, *Tetrahedron Lett.*, 25, 1379 (1984); H. Lönn, *Chem. Commun. Univ. Stockholm*, No2 (1984); Ref. 10a).
- 16) All values of $[\alpha]_D$ were measured for CHCl₃ solutions at 20°C. Compounds having $[\alpha]_D$ recorded gave satisfactory elemental analysis.
- 17) J.-R. Pougny, J.-C. Jacquinet, M. Nassr, D. Duchet, M.-L. Milat and P. Sinäy, J. Am. Chem. Soc., 99, 6762 (1977).
- 18) 9 α , [α]_D +33.6° (c 1.1), δ _H(CDCl₃) 5.728 (d, 3.9 Hz, H-1'), 4.543 (d, 3.7 Hz, H-1), δ _C(CDCl₃) 97.31 (C-1), 96.28 (C-1'); 9 β , [α]_D +37.2° (c 0.8), δ _H(CDCl₃), 4.752 (d, 7.8 Hz, H-1'), 4.613 (d, 3.7 Hz, H-1), δ _C(CDCl₃) 102.40 (C-1'), 98.27 (C-1).
- 19) 10α , $\delta_{H}(CDCl_{3})$ 5.671 (d, 3.7 Hz, H-1'), 4.551 (d, 7.8 Hz, H-1), $\delta_{C}(CDCl_{3})$ 102.35 (C-1), 96.71 (C-1'); 10β , $\delta_{H}(CDCl_{3})$ 4.511 (d, 8.1 Hz, H-1), 4.480 (d, 7.6 Hz, H-1'), $\delta_{C}(CDCl_{3})$ 102.51 (C-1'), 102.35 (C-1); Pure 10β , m.p. 96-98°C, $[\alpha]_{D}$ +8.1° (c 0.8) was isolated by recrystallization from n-hexane-diethyl ether.
- 20) Prepared from p-nitrobenzoate 12²¹) by the tributylstannyl method¹³). $[\alpha]_D$ +51.1° (c 1.6), $\delta_H(CDCl_3)$ 5.397 (dd, 9.8, 9.5 Hz, H-4), 5.264 (d, 1.7 Hz, H-1), 4.161 (ddd, 9.8, 6.1, 3.2 Hz, H-5), 3.812 (dd, 3.2, 1.7 Hz, H-2), 3.773 (dd, 9.4, 3.2 Hz, H-3), 3.625 (dd, 10.7, 6.1 Hz, H-6), 3.555 (dd, 10.7, 3.2 Hz, H-6), 2.123 and 1.926 (2s, OAc and SMe), $\delta_C(CDCl_3)$ 83.38 (164.8 Hz, C-1).
- 21) K. Koike, M. Mori, Y. Ito, Y. Nakahara and T. Ogawa, Glycoconjugate J., in press (1987).

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